

between the purine leaving group and the anionic nucleophile. This mechanism was first described for PNP and has since been shown to apply to other glycosyltransferases. With fixed nucleophiles in the catalytic site, a ribooxacarbenium-ion transition state is formed at some point during the ribosyl migration. The exact nature of PNP transition states is dictated by the migration distance and protein vibrational modes that form the transition state. Remarkable differences in transition-state structure can occur with nearly identical protein structures. These differences can guide the design of transition-state analogs that demonstrate specificity for enzyme variants from different species. Knowledge of the transition-state structures has permitted access to picomolar compounds with favorable pharmacokinetic properties. Two of these have entered clinical trials for T-cell disease. PNP has also served as one of the first enzymes for exploration of catalytic-site-induced substrate distortions by binding isotope effects. These results are providing new insights into the fundamental notions of enzymatic catalysis, induced fit, and transition-state structure.

REFERENCES

- Pauling, L. Molecular architecture and biological reactions. *Chem. Eng. News* **1946**, *24*, 1375–1377.
- Wolfenden, R. Transition state analogues for enzyme catalysis. *Nature* **1969**, *223*, 704–705.
- Wolfenden, R. Analog approaches to the structure of the transition state in enzyme reactions. *Acc. Chem. Res.* **1972**, *5*, 10–18.
- Jencks, W. Binding energy, specificity, and enzymic catalysis: the Circe effect. *Adv. Enzymol. Relat. Areas Mol. Biol.* **1975**, *43*, 219–410.
- Anderson, V. Ground state destabilization. In: *Encyclopedia of Life Sciences*. Chichester: John Wiley & Sons, Ltd.; **2001**, 1–5.
- Shih, I.; Been, M. Catalytic strategies of the hepatitis delta virus ribozymes. *Annu. Rev. Biochem.* **2002**, *71*, 887–917.
- Wu, N.; Mo, Y.; Gao, J.; Pai, E. Electrostatic stress in catalysis: structure and mechanism of the enzyme orotidine monophosphate decarboxylase. *Proc. Natl. Acad. Sci. U.S.A.* **2000**, *97*, 2017–2022.
- Amyes, T. L.; Wood, B. M.; Chan, K.; Gerlt, J. A.; Richard, J. P. Formation and stability of a vinyl carbanion at the active site of orotidine 5'-monophosphate decarboxylase: pKa of the C-6 proton of enzyme-bound UMP. *J. Am. Chem. Soc.* **2008**, *130*, 1574–1575.
- Bruice, T. C.; Lightstone, F. C. Ground state and transition state contributions to the rates of intramolecular and enzymatic reactions. *Acc. Chem. Res.* **1999**, *32*, 127–136.
- Lightstone, F. C.; Bruice, T. C. Ground state conformations and entropic and enthalpic factors in the efficiency of intramolecular and enzymatic reactions. 1. Cyclic anhydride formation by substituted glutarates, succinate, and 3,6-endoxo- Δ 4-tetrahydrophthalate monophenyl esters. *J. Am. Chem. Soc.* **1996**, *118*, 2595–2605.
- Antonioni, D.; Basner, J.; Núñez, S.; Schwartz, S. Computational and theoretical methods to explore the relation between enzyme dynamics and catalysis. *Chem. Rev.* **2006**, *106*, 3170–3187.
- Kohen, A.; Cannio, R.; Bartolucci, S.; Klinman, J. Enzyme dynamics and hydrogen tunnelling in a thermophilic alcohol dehydrogenase. *Nature* **1999**, *399*, 496–499.
- Agarwal, P.; Billeter, S.; Rajagopalan, P.; Benkovic, S.; Hammes-Schiffer, S. Network of coupled promoting motions in enzyme catalysis. *Proc. Natl. Acad. Sci. U.S.A.* **2002**, *99*, 2794–2799.
- Wong, K.; Selzer, T.; Benkovic, S.; Hammes-Schiffer, S. Impact of distal mutations on the network of coupled motions correlated to hydride transfer in dihydrofolate reductase. *Proc. Natl. Acad. Sci. U.S.A.* **2005**, *102*, 6807–6812.
- Nunez, S.; Antonioni, D.; Schramm, V. L.; Schwartz, S. D. Promoting vibrations in human purine nucleoside phosphorylase: a molecular dynamics and hybrid quantum mechanical/molecular mechanical study. *J. Am. Chem. Soc.* **2004**, *126*, 15720–15729.
- Saen-Oon, S.; Ghanem, M.; Schramm, V.; Schwartz, S. Remote mutations and active site dynamics correlate with catalytic properties of purine nucleoside phosphorylase. *Biophys. J.* **2008**, *94*(10), 4078–4088.
- Lienhard, G. Enzymatic catalysis and transition-state theory. *Science* **1973**, *180*, 149–154.
- Wolfenden, R.; Kati, W. M. Testing the limits of protein-ligand binding discrimination with transition-state analogue inhibitors. *Acc. Chem. Res.* **1991**, *24*, 209–215.
- Wolfenden, R. Conformational aspects of inhibitor design: enzyme-substrate interactions in the transition state. *Bioorg. Med. Chem.* **1999**, *7*, 647–652.
- Schramm, V. Enzymatic transition state theory and transition state analogue design. *J. Biol. Chem.* **2007**, *282*, 28297–28300.
- Schramm, V. Enzymatic transition states and transition state analog design. *Annu. Rev. Biochem.* **1998**, *67*, 693–720.
- Hammond, G. S. A correlation of reaction rates. *J. Am. Chem. Soc.* **1955**, *77*, 334–338.
- Jencks, W. In: *Catalysis in Chemistry and Enzymology*. Dover: New York, **1987**, 170–182.
- Schramm, V. Enzymatic transition states: thermodynamics, dynamics and analogue design. *Arch. Biochem. Biophys.* **2005**, *433*, 13–26.
- Cleland, W. Isotope Effects: Determination of enzyme transition state structure. *Methods Enzymol.* **1995**, *249*, 341–373.
- Parkin, D. W. In: *Enzyme Mechanism from Isotope Effects*, Cook, P. F.; Ed. Boca Raton: CRC Press; **1991**, 269–290.
- Rodgers, J.; Femec, D. A.; Schowen, R. L. Isotopic mapping of transition-state structural features associated with enzymic catalysis of methyl transfer. *J. Am. Chem. Soc.* **1982**, *104*, 3263–3268.
- Sunhel, J.; Schowen, R. In: *Enzyme Mechanism from Isotope Effects*, Cook, P. F.; Ed. Boca Raton: CRC Press; **1991**, 3–36.
- Huskey, W. In: *Enzyme Mechanism from Isotope Effects*, Cook, P. F.; Ed. Boca Raton: CRC Press; **1991**, 37–72.
- Berti, P. J.; Tanaka, K. S. E. Transition state analysis using multiple kinetic isotope effects: mechanisms of enzymatic and non-enzymatic glycoside hydrolysis and transfer. *Adv. Phys. Org. Chem.* **2002**, *37*, 239–314.
- Rose, I. The isotope trapping method: desorption rates of productive e.s complexes. *Methods Enzymol.* **1980**, *64*, 47–59.
- Frisch, M. J.; Trucks, G. W.; Schlegel, H. B.; Scuseria, G. E.; Robb, M. A.; Cheeseman, J. R.; Zakrzewski, V. G.; Montgomery, J.; Stratmann, R. E.; Burant, J. C.; Dapprich, S.; Millam, J. M.; Daniels, A. D.; Kudin, K. N.; Strain, M. C.; Farkas, O.; Tomasi, J.; Barone, V.; Cossi, M.; Cammi, R.; Mennucci, B.; Pomelli, C.; Adamo, C.; Ochterski, J.; Petersson, G. A.; Ayala,